

## THERAPEUTIC TARGETING OF MOLECULAR PATHWAYS IN COLORECTAL CANCER

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Mutations in tumor suppressor genes, cell signaling, and genes associated with DNA repair lead to onset of colorectal cancer (CRC). Even though most CRC patients get clinical benefits from conventional treatments such as chemotherapy and radiotherapy, treatment success is still not at the desired level despite recent advances in CRC treatments. Therefore, further elucidation of the molecular signaling pathways involved in CRC progression will allow developing targeted therapies. With the detection of signaling pathways that lead to cancer progression and development of the successful treatment methods targeting these pathways, the progression of the disease can be prevented. This review provides an overview of the therapeutic roles of potential molecular targets in recent preclinical and clinical studies in CRC treatment.

**Key Words:** colorectal cancer drugs, molecular targeted therapy, CRC signaling pathways, tumor biomarkers.

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Colorectal cancer (CRC) is the most common malignancy in the world. According to estimates by 2030, CRC incidence in the world is expected to increase by 60% with 2.2 million new cases and 1.1 million deaths [1]. Depending on the origin of the mutation, colorectal carcinomas are classified as sporadic and hereditary [2]. The most frequently occurring mutated oncogenes in CRC are *KRAS* and *c-MYC* [3]. *KRAS* is an oncogene responsible for the synthesis of a protein involved in the transmission of mitogenic messages in the cell membrane. Point mutations in *KRAS* are observed in 39–71% of CRC and 42% of adenomatous polyps [4–6].

The following molecular signaling pathways associated with genomic instability have been identified in CRC: chromosomal instability (CIN), microsatellite instability (MSI), and CpG island methylator phenotype pathways [7]. In addition to *KRAS* mutations in CRC, major mutations in the transforming growth factor- $\beta$  (TGF- $\beta$ ), PIK3CA and TP53 signaling also develop. Single colorectal tumor may contain up to 80 mutated genes [7–8]. These important molecular properties and cell signaling pathways could be the targets in personalized therapies. Moreover, the various strategies such as angiogenesis inhibitors, EGFR-targeted therapy, BRAF mutation-targeted therapies and immunotherapy may be used. In this context, it is quite important that effective and non-toxic targeted therapies based on molecular signaling be discovered [7].

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**Abbreviations used:** APC – adenomatous polyposis coli; CIN – chromosomal instability; CRC – colorectal cancer; EGFR – epidermal growth factor receptor; FZD – Wingless/Int1-Frizzled; MAPK – mitogen-activated protein kinase; MSI – microsatellite instability; PD-1 – programmed cell death-1; PORCN – porcupine-O-acetyltransferase; TGF- $\beta$  – transforming growth factor- $\beta$ ; TNKS – tankyrase; VEGF – vascular endothelial growth factor.

### THERAPEUTIC MOLECULAR TARGETING OF APC

In CRC, specific mutations that stimulate oncogenic signaling occur in CIN tumors (Table 1). One of the most important mutations that cause CIN is adenomatous polyposis coli (APC) gene mutations that occur in approximately 80% of CRC, while 5% to 10% have mutational changes in Wnt signaling [5, 9]. The APC gene is not only a crucial switch of the Wnt signaling mechanism but also plays a critical role in regulating chromosomal differentiation, cell differentiation, adhesion, migration, and apoptosis. Most general sporadic CRC exhibit unusual activation of the Wnt signaling. The genetic defect of APC leads to the activation of the Wnt signaling mechanism being an important initial genetic condition for CRC. APC mutations play an important role in the development of both familial adenomatous polyposis (FAP) and sporadic CRC. This resulting loss of function in the APC gene triggers a series of molecular-genetic instability and histological changes leading to malignant transformation [10]. As the therapeutic modality for APC targeting, a small molecule called Truncated APC Selective Inhibitor 1 has been shown to inhibit tumor growth in the CRC mouse model with minimal toxicity. The molecule exerts its cytotoxicity independent of Wnt pathway inhibition and does not specifically exert detrimental effects on normal mouse colonic epithelium [11]. Understanding the comprehensive functions of APC mutations will shed light on the molecular mechanisms of CRC tumorigenesis and uncover new drug targets that allow developing additional targeted therapeutics for CRC treatment.

### THERAPEUTIC MOLECULAR TARGETING OF WNT PATHWAY

Wnt signaling activation is critical in tumor progression. Molecular targets for Wnt signaling inhibition such as porcupine-O-acetyltransferase (PORCN), FZD (Wingless/Int1-Frizzled), Dvl,  $\beta$ -catenin degradation complex,

**Table 1.** Frequency of genetic mutation in CIN positive CRC [7]

| Gene                           | Chromosomal location | Frequency of mutations (%) | Role of a gene product?                                               |
|--------------------------------|----------------------|----------------------------|-----------------------------------------------------------------------|
| <b>Oncogenes</b>               |                      |                            |                                                                       |
| <i>KRAS</i>                    | 12p12                | ~30–50                     | Cell survival, proliferation and transformation                       |
| <i>BRAF</i>                    | 7q34                 | ~40–60                     | Cell growth, cell division, differentiation, and secretion            |
| <i>CTNNB1</i>                  | 3p22                 | ~4–15                      | Regulation of Wnt signaling target genes that drive tumor development |
| <i>PIK3CA</i>                  | 3q26                 | ~20                        | Cell survival, proliferation                                          |
| <b>Tumor suppressor genes</b>  |                      |                            |                                                                       |
| <i>APC</i>                     | 5q21                 | ~30–70                     | Inhibition of WNT pathway, cytoskeletal regulation                    |
| <i>TP53</i>                    | 17p13                | ~40–50                     | Apoptosis induction, cell cycle arrest                                |
| <i>SMAD4</i> ,<br><i>SMAD2</i> | 18q21                | ~10–20                     | Intracellular mediators of the TGF- $\beta$ signaling                 |
| <i>DCC</i>                     | 18q21                | ~6                         | Cell surface receptor for netrin-1                                    |

nuclear  $\beta$ -catenin, and tankyrase (TNKS) have been identified [11]. Monoclonal antibodies and small-molecule inhibitors have been identified as two therapeutic modalities for targeting Wnt signaling and demonstrated antitumor effects [12]. Several small-molecule inhibitor trials have been conducted in the preclinical and clinical phases of drug research studies, and  $\beta$ -catenin inhibitors PRI-724, CWP232228, and BC2059 that suppress TCF/LEF target genes have been developed [13]. TCF/LEF transcription factors, major endpoint mediators of Wnt/Wingless signaling, are multifunctional proteins that use sequence-specific DNA binding to determine which genes are regulated by Wnt [14]. In previous studies, it was revealed that tumor development was stopped by inhibiting the Wnt signaling pathway, together with the inhibition of the  $\beta$ -catenin/TCF interaction [15]. However, there are two TNKS inhibitors that attenuate Wnt signaling, XAV939 and E7449 [11]. WNT974 and ETC-159 are found as PORCN inhibitors [11]. The small-molecule inhibitor ETC-159 is in phase I clinical trials for the therapy of CRC, while WNT974 is in phase I/II for the treatment of metastatic CRC adenocarcinoma [16–18]. While blocking Wnt pathway via small-molecule inhibitors is a possible therapeutic approach to stop tumor development, further research is required to discover how these inhibitors may also influence other signaling mechanisms in order to decrease toxicity to normal cells [11].

Another way to regulate Wnt signaling is the application of monoclonal antibodies. There are several antibody-based therapies currently under study (Table 2). Phase I clinical trials are ongoing for OMP-18R5, monoclonal antibody, solid tumors targeting the five FZD receptors [19]. Finally, it is in phase I clinical trial for OMP-131R10, an anti-R-spondin 3 antibody for the treatment of solid tumors and metastatic CRC [16]. The application of monoclonal antibodies to target Wnt pathway is promising, but further clinical research is required to determine the clinical importance.

### THERAPEUTIC MOLECULAR TARGETING OF TGF- $\beta$ PATHWAY

In colorectal tumorigenesis, chromosomal changes involving TGF- $\beta$  participate in the CIN pathway. Loss

**Table 2.** Biomarkers of Wnt signaling [20] and Wnt-targeted monoclonal antibodies [21]

| Agents of Wnt signaling in CRC under clinical trial     |            |                         |                                           |
|---------------------------------------------------------|------------|-------------------------|-------------------------------------------|
| Molecular signaling mechanism                           | Agent      | Phase                   | NCT identifier                            |
| Wnt/ $\beta$ -catenin signaling                         | WNT-974    | Phase 1/2               | NCT02278133                               |
|                                                         | FOXY-5     | Phase 1                 | NCT02655952                               |
|                                                         | LGK-974    | Phase 1                 | NCT02020291<br>NCT02020291<br>NCT01351103 |
| Wnt-targeted monoclonal antibodies under clinical trial |            |                         |                                           |
| Target                                                  | Agent      | Phase                   | Ref.                                      |
| R-spondin3                                              | OMP-131R10 | Phase I                 | [15, 22]                                  |
| FZD10                                                   | OTSA101    | Phase 1<br>(terminated) | [15, 23]                                  |

of function of *SMAD2* and *SMAD4* tumor suppressor genes inactivates TGF- $\beta$  signaling mechanism, thereby increasing cell proliferation and escaping apoptosis [9]. In the majority of CRC, activation of MYC, which has an important role in colorectal carcinoma, occurs through inactivation of the TGF- $\beta$  signaling and/or activation of the Wnt molecular pathway [9].

In CRC, disruptions in TGF- $\beta$  signaling lead to metastasis and increased tumor growth [24]. For this reason, the TGF- $\beta$  pathway has become an important molecular pathway for drug development studies. Current candidate anti-TGF- $\beta$  therapies have focused on the targeting of TGF- $\beta$  signaling at the ligand-receptor level using monoclonal antibodies or peptides and the use of TGF- $\beta$  receptor kinase inhibitors [24]. However, the TGF- $\beta$  pathway is quite complex, with different effects in normal cells and different types of cancer, and paradoxical functions in the immune system modulation and tumor microenvironment. Although TGF- $\beta$  inhibition has not been used yet in the treatment of CRC, the studies of many candidate molecules are continuing. A list of current TGF- $\beta$  signaling inhibitors under research can be seen in Table 3 [24].

**Table 3.** TGF- $\beta$  receptor and ligand inhibitors under clinical investigation [24–25]

| Agent                            | Target          | NCT identifier | Status                            |
|----------------------------------|-----------------|----------------|-----------------------------------|
| TGF- $\beta$ ligand inhibitors   |                 |                |                                   |
| Trabedersen                      | TGF- $\beta$ 2  | NCT00844064    | Phase I study in CRC completed    |
| FANG vaccine                     | TGF- $\beta$ 1, | NCT01505166    | Phase II study in CRC ongoing     |
|                                  | TGF- $\beta$ 2  | NCT01453361    |                                   |
| TGF- $\beta$ receptor inhibitors |                 |                |                                   |
| PF-03446962                      | TGF- $\beta$ RI | NCT00557856    | Phase I study in CRC completed    |
|                                  |                 | NCT01620970    | Phase I study in combination with |
|                                  |                 | NCT02116894    | regorafenib in CRC completed      |

### THERAPEUTIC MOLECULAR TARGETING OF VEGF

Vascular endothelial growth factor (VEGF) is an angiogenic growth factor and the ability of VEGF signaling determines the endothelial cell survival, proliferation, and migration. One study demonstrated that bevacizumab, a VEGF-blocking monoclonal antibody, provided benefit in CRC upon addition to fluorouracil-based combination chemotherapy regimens [26]. Blocking VEGF-A with bevacizumab has proved hopeful in several phase III clinical experiments. Other strategies

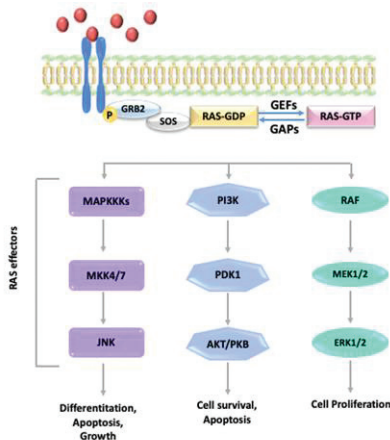
for targeting VEGF signaling involve the direct binding of monoclonal antibodies such as ramucirumab to the extracellular domain of the VEGF receptor (VEGFR-2) and the use of tyrosine kinase inhibitors for antiangiogenic effect [26] (Table 4).

**Table 4.** Agents of VEGFR in CRC under clinical trial

| Agent      | Target     | Phase   | NCT identifier |
|------------|------------|---------|----------------|
| LYN00101   | VEGF       | Phase 1 | NCT03644459    |
| Icrucumab  | VEGFR      | Phase 2 | NCT01111604    |
| Vandetanib | VEGFR/EGFR | Phase 2 | NCT00454116    |
| Pazopanib  | Pan-VEGFR  | Phase 1 | NCT00387387    |

**THERAPEUTIC MOLECULAR TARGETING OF RAS PATHWAY**

Epidermal growth factor receptor (EGFR), one of the transmembrane receptors, organizes many important cell functions including raising cell proliferation, cell cycle progression, and inhibiting apoptosis by the activation of intracellular signaling including phosphoinositide 3-kinase, mitogen-activated protein kinase (MAPK), and Signal Transducer and Activator of Transcription proteins [27–28]. Various mutations, overexpression or stimulation in MAPK and PI3K signaling pathways, which play an active role in cell proliferation, provide proliferative benefits for tumor cells. The most common mutations affecting MAPK/PI3K signaling in CRC are *KRAS*, *BRAF* and *PIK3CA* [29]. *KRAS* mutations occur in approximately 40% of CRCs [28, 30]. Especially, abnormal intracellular RAS signaling has various roles within the tumor microenvironment TME. Oncogenic RAS mutations happen in a constitutively initiated state whereby RAS proteins are no longer self-inhibited through normal GTPase activity. Approaches to control mutant RAS signaling require single or dual targeting of RAS itself, the RAS-RAF-MEK-ERK axis, the PI3K-AKT axis (Figure) [31].



**Figure.** RAS molecular signaling pathway in cancer. Growth factors binding to their cell surface receptors activate guanine exchange factors (GEF), such as SOS (son of sevenless) that are connected by the adaptor protein GRB2 (growth-factor-receptor bound protein 2). SOS stimulates the release of bound guanosine diphosphate (GDP) from RAS, and it is exchanged for guanosine triphosphate (GTP), pointing to the active RAS-GTP conformation. The guanosine triphosphatase (GTPase)-activating proteins (GAP) can bind to RAS-GTP and stimulate the transformation of RAS-GTP to RAS-GDP, which stops signaling. Mutated RAS is constitutively active in the RAS-GTP conformation. Activated RAS organizes various cellular functions through effectors including MAPK pathway, Raf–MEK–ERK signaling, PI3K molecular pathway

Monoclonal antibodies cetuximab and panitumumab target the EGFR providing an additional clinical therapeutic benefit for CRC patients [32]. Although anti-EGFR monoclonal antibody therapy is used in CRC, mutations in these signaling pathways are resistant to treatment [27]. As with *RAS* and *BRAF* mutations, *PIK3CA* mutations are thought to confer resistance to anti-EGFR treatment. Therefore the molecular properties of EGFR signaling should be investigated in more detail to reveal potential targets for more effective therapy modalities in CRC [29].

**THERAPEUTIC MOLECULAR TARGETING OF COX2**

Various studies have also shown that COX2 (PTGS2), a molecule produced by prostaglandins that perform a significant function in inflammatory regulation, is overexpressed in CRC cases with *PIK3CA* mutations. For instance, aspirin suppresses COX2 expression and PIK3 signaling pathway. Studies have shown that regular use of aspirin in CRC reduces the risk of overexpression of PTGS2 [33].

**THERAPEUTIC MOLECULAR TARGETING OF TP53**

TP53, a tumor suppressor gene, is also a cell cycle checkpoint regulatory [7]. With the inactivation of TP53, excessive cell proliferation occurs and tumor progression is ensured. In addition, studies have shown that the transition from adenoma to invasive carcinoma is generally with the inactivation of TP53 [5, 7, 34]. Loss of function of chromosome 17q location where TP53 is localized is a frequent occurrence in colorectal tumors because TP53 operates a significant function in adenocarcinoma [9]. Even though TP53 gene mutations in cancer treatment are still not fully understood, possible TP53-targeted therapies have been proposed both for wild-type and mutant TP53 [35–36]. TP53-based therapies can be grouped according to various treatment strategies. These include restoration of normal p53 function lost by genomic mutation or direct targeting of p53-deficient cells [37–39]. Therefore, molecular characteristics of TP53 gene should be described in detail as potential targets in CRC.

**THERAPEUTIC MOLECULAR TARGETING OF PD-1/PD-L1**

When PD-L1 (programmed cell death ligand-1) on the surface of tumor cells binds to the programmed cell death-1 (PD-1) receptor on T cells, the resulting inhibitory signal modulates cytotoxic T-cell responses, blocking the antitumor immune response and thus tumor cell death. Blocking PD-1 or PD-L1 with monoclonal antibodies has produced important clinical results in many different types of cancer, such as melanoma, bladder cancer. 60% response rate to the anti-PD-L1 inhibitor pembrolizumab was determined in different CRCs [40]. There are candidate PD-1/PD-L1 molecules whose clinical studies are still ongoing for use in the treatment of CRC. A list of current ap-

proved immunotherapeutics can be seen in Tables 5, 6 below. The use of monoclonal antibodies to target PD-1/PD-L1 is promising, but further clinical research is required to discover clinical importance.

**Table 5.** Targeted CRC immunotherapy drugs [25]

| Approved drug   | Target                                          | Reviews of phase III clinical trials leading to approval |
|-----------------|-------------------------------------------------|----------------------------------------------------------|
| Panitumumab     | Wild-type KRAS and NRAS                         | Hocking et al. [41]                                      |
| Cetuximab       | Wild-type KRAS and NRAS                         | Hoyle et al. [42]                                        |
| Bevacizumab     | Vascular endothelial growth factor (VEGF)       | Ilic et al. [43]                                         |
| Ramucirumab     | VEGFR2                                          | Verdaguer et al. [44]                                    |
| Regorafenib     | Dual targeted VEGFR2-tyrosine kinase inhibition | Yoshino et al. [45]                                      |
| Ziv-Aflibercept | VEGF                                            | Perkins et al. [46]                                      |
| Pembrolizumab   | MSI-H or dMMR                                   | n/a                                                      |
| Nivolumab       | PD-1                                            | Rajan et al. [47]                                        |

**Table 6.** Immune checkpoint modulators in CRC under clinical trial [48]

| Agent        | Target      | Phase     | NCT identifier |
|--------------|-------------|-----------|----------------|
| Atezolizumab | PD-1/PD-L1  | Phase 3   | NCT02912559    |
| JS-001       | Toripalimab | Phase 1–2 | NCT03946917    |
|              |             | Phase 2   | NCT04118933    |
| AMP-224      | PD-1/PD-L1  | Phase 1   | NCT02298946    |
| TSR-033      | PD-1/PD-L1  | Phase 1   | NCT03250832    |
| Tremelimumab | CTLA-4      | Phase 1/2 | NCT03206073    |

Potential advances in determining the molecular characteristics of CRC will allow finding therapeutic targets for molecularly characterized subtypes. Furthermore, identifying important molecular signaling mechanisms for a particular CRC subtype may provide possible therapeutic strategies that enable better therapeutic approaches. Moreover, cancer immunotherapy is developing as a hopeful strategy for the treatment of various malignancies, including CRC. The complex biological heterogeneity of tumors complicates the implementation of successful treatment. Therefore, it is possible to discover effective molecular biomarkers for CRC and to determine cell signaling mechanisms. Thus, the prognosis of the disease and treatment responses will be better managed, and the potential for personalized treatment will increase [49–50].

In conclusion, although many aspects of molecular classifications of CRC have been identified as potential targets for personalized therapies or prognostic predictors, more information is needed for successful targeted therapy and prognostic procedures in routine clinical application.

## REFERENCES

1. Arnold M, Sierra MS, Laversanne M, *et al.* Global patterns and trends in colorectal cancer incidence and mortality. *Gut* 2017; **66**: 683–91. doi:10.1136/gutjnl-2015-310912
2. Mármol I, Sánchez-de-Diego C, Dieste AP, *et al.* Colorectal carcinoma: A general overview and future perspectives in colorectal cancer. *Int J Mol Sci* 2017; **18**. doi:10.3390/ijms18010197
3. Meltzer SJ, Ahnen DJ, Battifora H, *et al.* Protooncogene abnormalities in colon cancers and adenomatous polyps. *Gastroenterology* 1987; **92**: 1174–80. doi:10.1016/s0016-5085(87)91074-2
4. Forrester K, Almoguera C, Han K, *et al.* Detection of high incidence of K-ras oncogenes during human colon tumorigenesis. *Nature* 1987; **327**: 298–303. doi:10.1038/327298a0
5. Vogelstein B, Fearon ER, Hamilton SR, *et al.* Genetic alterations during colorectal-tumor development. *New England J Med* 1988; **319**: 525–32. doi:10.1056/NEJM198809013190901
6. Losi L, Benhattar J, Costa J. Stability of K-ras mutations throughout the natural history of human colorectal cancer. *Eur J Cancer* 1992; **28A**: 1115–20. doi:10.1016/0959-8049(92)90468-h
7. Pino MS, Chung DC. The chromosomal instability pathway in colon cancer. *Gastroenterology* 2010; **138**: 2059–72. doi:10.1053/j.gastro.2009.12.065
8. Wood LD, Parsons DW, Jones S, *et al.* The genomic landscapes of human breast and colorectal cancers. *Science* 2007; **318**: 1108–13. doi:10.1126/SCIENCE.1145720
9. Cancer Genome Atlas Network. Comprehensive molecular characterization of human colon and rectal cancer. *Nature* 2012; **487**: 330–7. doi:10.1038/nature11252
10. Fodde R. The APC gene in colorectal cancer. *Eur J Cancer* 2002; **38**: 867–71. doi:10.1016/S0959-8049(02)00040-0
11. Tran FH, Zheng JJ. Modulating the wnt signaling pathway with small molecules. *Protein Sci* 2017; **26**: 650–61. doi:10.1002/PRO.3122
12. Anastas JN, Moon RT. WNT signalling pathways as therapeutic targets in cancer. *Nat Rev Cancer* 2013; **13**: 11–26. doi:10.1038/NRC3419
13. Katoh M, Katoh M. Molecular genetics and targeted therapy of WNT-related human diseases (Review). *Int J Mol Med* 2017; **40**: 587–606. doi:10.3892/IJMM.2017.3071
14. Cadigan KM, Waterman ML. TCF/LEFs and Wnt signaling in the nucleus. *Cold Spring Harb Perspect Biol* 2012; **4**: a007906. doi: 10.1101/cshperspect.a007906
15. Polakis P. Wnt signaling in cancer. *Cold Spring Harb Persp Biol* 2012; **4**: 9. doi:10.1101/CSHPERSPECT.A008052
16. Zhan T, Rindtorff N, Boutros M. Wnt signaling in cancer. *Oncogene* 2017; **36**: 1461–73. doi:10.1038/ONC.2016.304
17. Blagodatski A, Poteryaev D, Katanaev VL. Targeting the Wnt pathways for therapies. *Mol Cell Ther* 2014; **2**: 28. doi:10.1186/2052-8426-2-28
18. Zhang X, Hao J. Development of anticancer agents targeting the Wnt/ $\beta$ -catenin signaling. *Am J Cancer Res* 2015; **5**: 2344–60.
19. Katoh M. Canonical and non-canonical WNT signaling in cancer stem cells and their niches: Cellular heterogeneity, omics reprogramming, targeted therapy and tumor plasticity (Review). *Int J Oncol* 2017; **51**: 1357–69. doi:10.3892/IJO.2017.4129
20. Prosperi JR, Luu HH, Goss KH. Dysregulation of the Wnt pathway in solid tumors. In: Goss KH, Kahn M, eds. *Targeting the Wnt Pathway in Cancer*. Springer, 2011: 81–128. doi:10.1007/978-1-4419-8023-6\_5
21. Richman SD, Jasani B. Predictive biomarkers and targeted therapies in colorectal cancer. In: Badve S, Kumar GL, eds. *Predictive Biomarkers in Oncology*. Springer Nature Switzerland, 2019: 423–30. doi:10.1007/978-3-319-95228-4\_38
22. Mohammed MK, Shao C, Wang J, *et al.* Wnt/ $\beta$ -catenin signaling plays an ever-expanding role in stem cell self-renewal, tumorigenesis and cancer chemoresistance. *Genes Dis* 2016; **3**: 11–40. doi:10.1016/J.GENDIS.2015.12.004
23. de Lau W, Peng WC, Gros P, Clevers H. The Rspodin/Lgr5/Rnf43 module: regulator of Wnt signal strength. *Genes Dev* 2014; **28**: 305–16. doi:10.1101/GAD.235473.113
24. Neuzillet C, Tijeras-Raballand A, Cohen R, *et al.* Targeting the TGF $\beta$  pathway for cancer therapy. *Pharm Ther* 2015; **147**: 22–31. doi:10.1016/J.PHARMTHERA.2014.11.001
25. Badve S, Kumar GL. *Predictive Biomarkers in Oncology*. Springer Nature Switzerland, 2019. <https://link.springer.com/book/10.1007/978-3-319-95228-4>

26. Hurwitz H, Fehrenbacher L, Novotny W, *et al.* Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. *New Engl J Med* 2004; **350**: 2335–42. doi:10.1056/NEJM0A032691
27. De Roock W, Claes B, Bernasconi D, *et al.* Effects of KRAS, BRAF, NRAS, and PIK3CA mutations on the efficacy of cetuximab plus chemotherapy in chemotherapy-refractory metastatic colorectal cancer: A retrospective consortium analysis. *Lancet Oncol* 2010; **11**: 753–62. doi:10.1016/S1470-2045(10)70130-3
28. Lee D-W, Han S-W, Cha Y, *et al.* Association between mutations of critical pathway genes and survival outcomes according to the tumor location in colorectal cancer. *Cancer* 2017; **123**: 3513–23. doi:10.1002/cncr.30760
29. Dienstmann R, Connor K, Byrne AT; COLOSUS Consortium. Precision therapy in RAS mutant colorectal cancer. *Gastroenterology* 2020; **158**: 806–11. doi:10.1053/J.GASTRO.2019.12.051
30. Inamura K, Yamauchi M, Nishihara R, *et al.* Prognostic significance and molecular features of signet-ring cell and mucinous components in colorectal carcinoma. *Ann Surg Oncol* 2015; **22**: 1226–35. doi:10.1245/s10434-014-4159-7
31. Jones RP, Sutton PA, Evans JP, *et al.* Specific mutations in KRAS codon 12 are associated with worse overall survival in patients with advanced and recurrent colorectal cancer. *Br J Cancer* 2017; **116**: 923–9. doi:10.1038/bjc.2017.37
32. Cunningham D, Humblet Y, Siena S, *et al.* Cetuximab monotherapy and cetuximab plus irinotecan in irinotecan-refractory metastatic colorectal cancer. *New Engl J Med* 2004; **351**: 337–45. doi:10.1056/NEJM0A033025
33. Chan AT, Ogino S, Fuchs CS. Aspirin use and survival after diagnosis of colorectal cancer. *JAMA* 2009; **302**: 649–58. doi:10.1001/jama.2009.1112
34. Lengauer C, Kinzler KW, Vogelstein B. Genetic instabilities in human cancers. *Nature* 1998; **396**: 643–9. doi:10.1038/25292
35. Hoadley KA, Yau C, Wolf DM, *et al.* Multiplatform analysis of 12 cancer types reveals molecular classification within and across tissues of origin. *Cell* 2014; **158**: 929–44. doi:10.1016/J.CELL.2014.06.049
36. Vogelstein B, Kinzler KW. Achilles' heel of cancer? *Nature* 2001; **412**: 865–6. doi:10.1038/35091170
37. Roth JA, Grammer SF, Swisher SG, *et al.* Gene replacement strategies for treating non-small cell lung cancer. *Semin Radiat Oncol* 2000; **10**: 333–42. doi:10.1053/SRAO.2000.9127
38. Bischoff JR, Kirn DH, Williams A, *et al.* An adenovirus mutant that replicates selectively in p53-deficient human tumor cells. *Science* 1996; **274**: 373–6. doi:10.1126/science.274.5286.373
39. Tanaka T, Watanabe M, Yamashita K. Potential therapeutic targets of TP53 gene in the context of its classically canonical functions and its latest non-canonical functions in human cancer. *Oncotarget* 2018; **9**: 16234–47. doi:10.18632/oncotarget.24611
40. Le DT, Durham JN, Smith KN, *et al.* Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. *Science* 2017; **357**: 409–13. doi:10.1126/science.aan6733
41. Hocking CM, Price TJ. Panitumumab in the management of patients with KRAS wild-type metastatic colorectal cancer. *Ther Adv Gastroenterol* 2014; **7**: 20. doi:10.1177/1756283X13498660
42. Hoyle M, Crathorne L, Peters J, *et al.* The clinical effectiveness and cost-effectiveness of cetuximab (mono- or combination chemotherapy), bevacizumab (combination with non-oxaliplatin chemotherapy) and panitumumab (monotherapy) for the treatment of metastatic colorectal cancer after first-line chemotherapy (review of technology appraisal no. 150 and part review of technology appraisal no. 118): A systematic review and economic model. *Health Technol Assess* 2013; **17**: 1–237. doi:10.3310/HTA17140
43. Ilic I, Jankovic S, Ilic M. Bevacizumab combined with chemotherapy improves survival for patients with metastatic colorectal cancer: evidence from meta analysis. *PloS One* 2016; **11**: e0161912. doi:10.1371/JOURNAL.PONE.0161912
44. Verdaguer H, Tabernero J, Macarulla T. Ramucirumab in metastatic colorectal cancer: evidence to date and place in therapy. *Ther Adv Med Oncol* 2016; **8**: 230. doi:10.1177/1758834016635888
45. Yoshino T, Komatsu Y, Yamada Y, *et al.* Randomized phase III trial of regorafenib in metastatic colorectal cancer: analysis of the CORRECT Japanese and non-Japanese subpopulations. *Invest New Drugs* 2015; **33**: 740–50. doi:10.1007/S10637-014-0154-X
46. Perkins SL, Cole SW. Ziv-aflibercept (Zaltrap) for the treatment of metastatic colorectal cancer. *Ann Pharmacother* 2014; **48**: 93–8. doi:10.1177/1060028013506562
47. Rajan A, Kim C, Heery CR, *et al.* Nivolumab, anti-programmed death-1 (PD-1) monoclonal antibody immunotherapy: Role in advanced cancers. *Human Vaccines Immunother* 2016; **12**: 2219. doi: 10.1080/21645515.2016.1175694
48. Xie Y-H, Chen Y-X, Fang J-Y. Comprehensive review of targeted therapy for colorectal cancer. *Signal Transduct Target Ther* 2020; **5**: 1–30. doi: 10.1038/s41392-020-0116-z.
49. Inamura K. Colorectal cancers: An update on their molecular pathology. *Cancers* 2018; **10**: 26. doi:10.3390/cancers10010026
50. Alwers E, Jia M, Kloor M, *et al.* Associations between molecular classifications of colorectal cancer and patient survival: a systematic review. *Clin Gastroenterol Hepatol* 2019; **17**: 402–10. e2. doi:10.1016/j.cgh.2017.12.038

#### ТЕРАПЕВТИЧНІ МІШЕНІ МОЛЕКУЛЯРНИХ СИГНАЛЬНИХ ШЛЯХІВ У КЛІТИНАХ КОЛОРЕКТАЛЬНОГО РАКУ

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У виникненні злоякісних пухлин, у тому числі колоректального раку, задіяні мутації генів — супресорів пухлинного росту, генів білків сигнальних каскадів та генів, асоційованих з репараціями ДНК. Незважаючи на позитивні клінічні результати застосування хіміотерапії та променевої терапії у хворих на колоректальний рак, результати лікування ще далеко не досягають бажаного рівня. З'ясування молекулярних сигнальних шляхів, задіяних у прогресуванні захворювання, може сприяти розробці нових методів таргетної терапії. В огляді розглянуто потенційні молекулярні мішені, які вивчаються останнім часом у доклінічних та клінічних дослідженнях та можуть знайти своє застосування у лікуванні хворих на колоректальний рак.

**Ключові слова:** колоректальний рак, молекулярна таргетна терапія, сигнальні шляхи, пухлинні біомаркери.